

Rhizosphere *Streptomyces* Producing Antifungal to *Candida albicans* ATCC 10231 and *C. albicans* ATCC 24433 from Various Plants

Eti Sumiati¹, Dewi Nur Anggraeni²

¹Department of Health, Sekolah Tinggi Ilmu Kesehatan Mataram, Indonesia. 83116

²Faculty of Biology, Universitas Medan Area, Indonesia. 20223

{sumiatie070@gmail.com¹, dewinur@staff.uma.ac.id²}

Abstract. A research on potency of rhizosphere *streptomyces* producing antifungal on various plants (*Cyperus rotundus* L., *Zea mays* L., and *Rhizopora mucronanata*) was conducted. Twelve isolates of *streptomyces* was tested to assess their potency in producing antifungal compound by using agar block method with *Candida albicans* ATCC 10231 and *C. albicans* ATCC 24433 as test fungi. The result showed that 3 isolates among 12 isolates showed their potency to produce antifungal. Based on their antifungal activity, 1 isolates (J10) were chosen to produce antifungal compound using fermentation method in GYE broth. Characterization of antifungal compound was carried out by using TLC, GC-MS and bioautographic test. The result of bioautography test showed that the spot from TLC of J10 isolates could inhibit *C. albicans* ATCC 10231 growth. Identification of antifungal compound of J10 by using TLC analysis and spray reagent suggested that then alkaloid, flavonoid, and phenol class of compound.

Keywords: *Streptomyces*, *Rhizosphere*, *Antibiotic antifungal*.

1 Introduction

The emergence of the problem of infection caused by microbes, especially fungi in human life is a problem that does not end because it always raises new problems that are quite disturbing and detrimental to humans. These losses occur in various fields such as health (Akhand et al., 2010; Muiru et al., 2008), agriculture, and plantations (Joo, 2005; Boruwa et al., 2004) so that handling these problems is very necessary. Strains to the *Streptomyces* genus are microbial groups whose members are the largest antibiotic producers because 75% of the antibiotics that have been found are from strains to the genus *Streptomyces* (Akhand et al, 2010). Iwamoto et al. (1990) succeeded in finding a new (anticandide) antifungal FR109615 derived from *Streptomyces setonii* No. 7562. Ohkuma et al. (1992) also found Sultricin compounds produced by *S. roseiscleroticus* L827-7 (ATCC53903) which are new antifungal antibiotics as well as antitumor antibiotics. Furthermore Ueki et al. (1997) found UK-3A antifungal antibiotics produced by *Streptomyces* sp 517-02. Exploration and screening of secondary metabolites produced by strains *Streptomyces* are very important to obtain new compounds (antibiotics) that have the potential to inhibit harmful microbes (Hayakawa et al., 1996; Anupama et al., 2007; Singh et al., 2008).

The study of de Araujo et al. (2000) succeeded in finding members of the genus *Streptomyces* endophytes in corn roots and leaves which acted as antibacterial and antifungal producers. Rahayu et al. (2007) succeeded in obtaining 5 isolates of *Streptomyces* which had the potential to produce antibiotics from the rhizosphere of the scrub (*Crotalaria striata*), king grass (*Zoysia matrella* (L.) Merr) and jukut domdoman (*Chrysopogon aciculatus* (Retz) Trin). Based on this information, it is known that the opportunity to obtain bioactive compounds from *Streptomyces* genus members associated with the rhizosphere region is still quite large so that it is necessary to explore strains of *Streptomyces* from several rhizospheres to obtain antifungal compounds (Schlatter et al., 2009; Chatujinda et al., 2007; Balouiri et al., 2015). Thus, the purpose of this study was to determine the ability of *Streptomyces* isolates from various rhizosphere plants as antifungal producers and to determine the class of antifungal compounds produced by *Streptomyces* isolates.

2 Material and Methods

Twelve isolates of *Streptomyces* were namely R6, R7, R10, R18, NR1, NR4, NR20, NJ20, NJ25, J10, J16, and J20. Test fungi were *Candida albicans* ATCC 10231 and *Candida albicans* ATCC 24433. Medium for *Streptomyces* (SCA), for test fungi (YMEA), and fermentation medium (GYE). 2% Tween 80 solution, aquades, and material for identification of active compounds.

Screening of Antifungal-Producing *Streptomyces* Isolates; One-week-old *Streptomyces* isolates in oblique SCA medium suspended in 10 ml 2% Tween 80. The suspension was taken as much as 0.1 ml and inoculated on the SCA medium so that the spread plate was then incubated for two weeks. Subsequent culture results were used to test antifungal activity with a method so that blocks using cork borer were 12 mm in diameter. The test fungi that were incubated for two days were suspended in 10 ml of sterile 2% Tween 80. 0.1 ml of the test function was inoculated on a spread plate YMEA (yeast) medium. The inoculation results were perforated using cork borer with a diameter of 12 mm. The part that has been perforated is filled with agar block containing *Streptomyces* isolates. The medium then incubated at room temperature (Nedialkova & Naidenova, 2005) and observed the presence of inhibitory zones around the block.

Test for Selected Isolates; The best (selected) isolate fermented into liquid Glycerol Yeast Extract (GYE) medium. Preparation of inoculum, isolates aged 2 weeks were suspended into 2% Tween, 2 ml of suspension was inoculated into 40 ml liquid GYE medium. The medium was incubated in a shaker incubator at a temperature of 25°C at a speed of 150 rpm for two days. The inoculum that was made was taken as much as 5 ml and inoculated into a 100 ml liquid fermentation (GYE) medium, then incubated in a shaker incubator temperature of 25°C at a speed of 150 rpm for five days. The fermented product is then centrifuged at a speed of 5000 rpm for 20 minutes (Worang, 2003). The supernatant was taken and extracted with technical ethyl acetate using a separating funnel. The upper phase is taken and the lower phase is collected to be extracted again with ethyl acetate solvents up to three repetitions. The extraction results were evaporated at the waterbath temperature of 50°C to dry (dry extract).

Characterization and Identification of Antifungal Compounds; Monitoring compounds by TLC (a) The extract was bottled on a silica gel F254 plate using a capillary tube, with a 1 cm bottling distance between each extract, the silica plate was dried. The silica plate that has been bottled with the extract is put into the developer tank which has been filled with the mobile phase namely chloroform: ethyl acetate with a ratio of 10: 1. The chromatography results are air dried and seen under UV254 nm and UV366 nm and sprayed with spray reagents.

Bioautography Test (b) The silica gel plate produced by TLC or which has been developed in the developer tank is drained until the eluent evaporates (dries). Then the silica gel plate was cut along the eluent development boundary and then tested on the test fungi by attaching the plate to the test medium which was inoculated with the test fungi and stored in the refrigerator for 20 minutes or until the compound could be distributed into the medium containing the test function. After 20 minutes, the medium is removed from the refrigerator and the plate that has been attached to the medium is removed from the medium. The medium was incubated for 24 hours at room temperature and it was seen that the barrier area around the site was attached to the TLC result plate (Isnaeni, 2005). Identification of Active Compounds (c) Identification of active compounds was carried out by spraying the results of TLC with various spray reagents and by the GC-MS method.

3 Results and Discussion

1. Screening of Antifungal-Producing Streptomyces Isolates

Twelve streptomyces isolates were tested for their ability to inhibit the test fungi (pathogenic fungi). The isolates were tested for antifungal activity against *Candida albicans* ATCC 10231 and *C. albicans* ATCC 24433. The results showed that as many as 3 streptomyces isolates had the potential to produce antifungal compounds with different abilities in inhibiting the test function. These differences may be influenced by several factors, namely the composition of the medium and culture conditions such as pH, temperature, carbon source, nitrogen source, and different incubation times for each strain. streptomyces (Oskay, 2010). The screening of antifungal activity of each strain is very important given the presence of streptomyces strains capable of producing more than one type of secondary metabolite such as *S. griseus* which produces antibacterial streptomycin and cycloheximide antifungal (Whiffen et al., 1946).

The screening results of antifungal-producing streptomyces isolates are presented in Table 1. Of the three isolates, one isolate, J10, was considered to be the most effective in inhibiting the test fungus so that this isolate was selected for the production of antifungal compounds. Figure 1 is the result of antifungal activity of J10 isolates. Selected isolates, namely J10, were tested for the ability to produce antifungal compounds by fermentation using liquid GYE medium. Fermentation is carried out for five days resulting in a medium that is cloudy yellow (Image not shown).

Table 1. Results of screening for antifungal-producing streptomyces isolates

No	Test Isolates	Test Fungi	
		C.a 1	C.a 2
1	R6	0	2,3
2	R7	0	0
3	R10	0	0
4	R18	0	0
5	NR1	0	0
6	NR4	0	0
7	NR20	0	0
8	NJ20	0	0
9	NJ25	0	0
10	J10	5,2	5,5
11	J16	2,8	3,22
12	J20	0	0

13	41X	0	0
14	20	0	0
15	11	0	0
16	26	0	0
17	L10	0	0

Note: *Candida albicans* ATCC 10231 (C.a 1) dan *Candida albicans* ATCC 24433 (C.a 2)

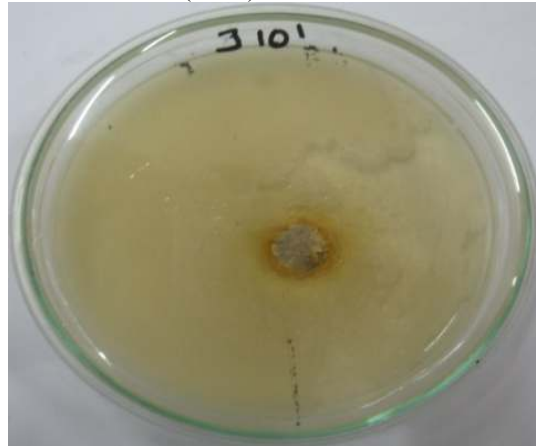


Figure 1. Anticandide activity of J10 isolates against *Candida albicans* ATCC 10231

2. Characterization and Identification of Antifungal Compounds

Three stages were carried out to characterize and identify antifungal compounds namely TLC monitoring compounds, bioautography tests, and identification with spray reagents and identification approaches using the GC-MS method. The results of monitoring compounds by TLC are presented in Figure 2, these results indicate that there are several spots detected. Detection of TLC on UV254 nm produces blue while UV366 nm produces purple fluorescence. The results of spraying with AlCl₃ give a yellow color that refers to flavonoid compounds, with purple anisaldehyde which refers to phenol compounds, and with dragendorff it produces a yellowish orange color indicating alkaloid compounds, with positive purple anisaldehyde for phenol, and positive brown cerium sulfate for general organic compounds that are matched based on Krebs et al. (1969) and Sutrisno (1986).

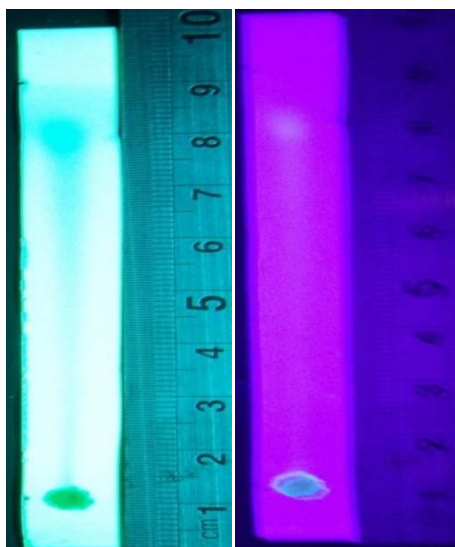


Figure 2. Results of TLC of ethyl acetate extract isolates J10 which detected UV254 (blue) and UV366 (purple)

The results of identification of compounds with several spray reactions showed that the ethyl acetate extract of J10 isolates contained flavonoid compounds, phenols and alkaloids based on the color produced after spraying with several spray reagents. Most of these compounds have been known to have antifungal activity. According to Cowan (1999) flavonoid compounds have activity to damage cell membranes and cell walls of fungi and can provide it with a fungi protein complex. While alkaloid compounds have the ability to bind to the DNA of fungi. It is possible that the activity of these two compounds causes inhibition and even death in fungal cells. The results of identification of compound groups using spray reagents are presented in Table 2.

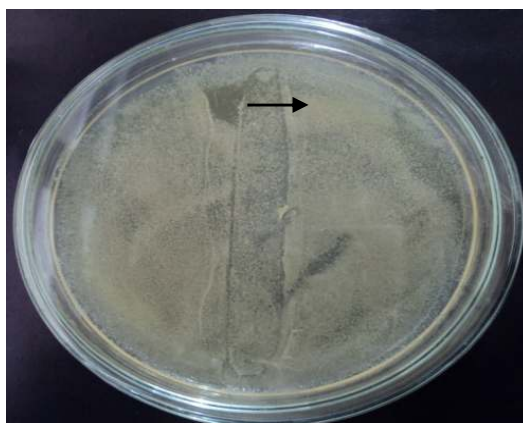


Figure 3. Results of bioautography test of J10 isolates against *Candida albicans* ATCC 10231

The bioautography test of ethyl acetate extract isolate J10 (TLC results) on *C. albicans* ATCC 10231 and ethyl acetate extract of J10 isolate are presented in Figure 3. The results showed that spot yield of TLC extract of ethyl acetate J10 isolate could inhibit *C. albicans* ATCC 10231.

Table 2. Results of identification of compound groups using TLC analysis and spray reagents.

No	Class of Compound	Spray Reagent	Standard Color	Result of Test	Rf J10
1	Flavonoids	AlCl ₃	Yellow	+	
2	Sugar	Anisaldehyde	0, 04		
3	Steroids	Anisaldehyde	Red	-	-
4	Terpen	Anisaldehyde	Green	-	-
5	Phenol	Anisaldehyde	Blue	-	-
6	Alkaloids	Dragendorff	Purple	+	
7	Amine	Ninhydrin	0,37&0,9		
8	Phenol	FeCl ₃	Yellow-orange	+	0,3
9	Hydroxamic acid	FeCl ₃	Blue	-	-
10	General Organic compound	Serium (IV) Sulfat	Blue Red	-	-
			Choco	+	0,5

Note: (+) Indicates the identification of a class of compounds

(-) Group of compounds not identified.

The results of identification of antifungal compounds by GC-MS showed that the ethyl acetate extract of J10 isolate was not detected by GC-MS. This is probably due to improper column selection and temperature regulation so that the compounds in the ethyl acetate extract J10 cannot be detected by GC-MS devices (GC-MS results are not shown). The GC-MS tool used in this study had a temperature of 300°C.

4 Conclusion

Streptomycetes isolates from various plant rhizosphere have the potential to produce antifungal compounds. The group of antifungal compounds produced by streptomycetes is a class of alkaloid compounds, phenols and flavonoids.

References

- [1] Akhand, M. A. M., Bari, M. A. A., Islam, M. A., and Khondkar1, P. 2010. Characterization and Antimicrobial Activities of a Metabolite from a New *Streptomyces* Species from Bangladeshi Soil. *Journal of Scientific Research* 2 (1): 178-185.
- [2] Anupama, M. Narayana, K. J. P., and Vijayalakshmi, M. 2007. Screening of *Streptomyces purpeofuscus* for Antimicrobial Metabolites. *Research Journal of Microbiology*. 2 (12): 992-994.
- [3] Balouiri, M., Bouhdid, S., Harki, H., E., Sadiki, M., Edrhiri, W., O., & Ibsouda, K., S., 2015. Antifungal activity Of Bacillus Spp. Isolates From Calotropis Procera

- Ait Rhizosphere against *Candida albicans*. *Asian Journal of Pharmaceutical and Clinical Research*. **8** (2)
- [4] Boruwa, J., Kalita, B., Barua, N. C., Borah, J. C., Mazumder, S., Thakur, D., Gogoi, D.K. & Bora, T. C., 2004. Synthesis, absolute stereochemistry and molecular design of the new antifungal and antibacterial antibiotic produced by *Streptomyces* sp.201. *Bioorganic & Medicinal Chemistry Letters*. **14** : 3571-3574.
- [5] Chatujinda, S., Tanasupawat, S., Amnuoypol, S., Chaichantipyuth, C., 2007. Identification And Antimicrobial Activities of *Streptomyces* Strain Isolated From Soils. *J Health Res* **21** (3): 195-200
- [6] Cowan, M. M., 1999, Plant Products as Antimicrobial Agents, *American Society for Microbiology*, Reviews. **12** (4); 564-582.
- [7] de Araujo, J. M., Silva, A. C. da., & Azevedo, J. L. 2000. Isolation of Endophytic Actinomycetes from Roots and leaves of Maize (*Zea mays*. L.) *Brazilian Archives of Biology and Technology*., **43** (4) : 52-58.
- [8] Hayakawa, M. Momose, Y, Yamazaki, T & Nonomura, H. 1996. A Method for the selective Isolation of Microtetraspora Glauca and Related Fourspered Actinomycetes from Soil. *Journal of Applied Bacteriol*. **80**: 375-386.
- [9] Isnaeni. 2005. Bioautografi Antibiotik Hasil Fermentasi Mutan *Streptomyces griseus* ATCC 10137. *Majalah Farmasi Airlangga*, **5** (1) : 16-19.
- [10] Iwamoto, T., Tsujii, E., Ezaki, M., Fujie, A., Hashimoto, S., Okuhara, M., Kohsaka, M., & Imanaka, H. 1990. FR109615, A New Antifungal Antibiotic from *Streptomyces setonii*. *Journal of Antibiotics*, **43** (1) : 1-7.
- [11] Joo, G. J. 2005. Production of An Antifungal Substance for Biological Control of *Phytophthora capsici* Causing Phytophthora Blight in Red-peppers by *Streptomyces halstedii*. *Biotechnology Letter*. **27** : 201-205.
- [12] Krebs, K.G., D. Heusser & H. Wimmer. Spray Reagents. In *Thin-Layer Chromatography : A Laboratory Handbook 2nd Edition* (E. Stahl Ed.). Springer-Verlag : Berlin.
- [13] Muiru, W. M., Mutitu, E. W., & Mukunya, D. M. 2008. Identification of Selected Actinomycete Isolates and Characterization of Their Antibiotic Metabolites. *Journal of Biological Sciences*. **8** (6): 1021-1026.
- [14] Nedialkova, D. & M. Naidenova. 2005. Screening The Antimicrobial Activity of Actinomycetes Strains Isolated from Antarctica. *Journal of Culture Collection* **4** : 29 -35.
- [15] Ohkuma, H., Naruse, N., Nishiyama, Y., Tsuno, T., Hoshino, Y., Sawada, Y., Konishi, M., and Oki, T., 1992. Sultricin a new Antifungal and Antitumor Antibiotic from *Streptomyces roseiscleroticus* Production, Isolation, Structure and Biological Activity. *The Journal of Antibiotics*. **45** (8): 1239-1249.
- [16] Oskay, M. 2009. Antifungal and Antibacterial Compounds from *Streptomyces* Strains. *African Journal of Biotechnology*. **8** (13) : 3007-3017.
- [17] Rahayu, T., Maryati, Sembiring, L., & Soegihardjo, C. J. 2007. Isolation and Characterization of *Streptomyces* potential as an Antimicrobial from Rizosphere. Collection of Summary of Research Results Workshop on Presentation of Research by DP2M Dikti in 2007. UMS, Surakarta.
- [18] Singh, V., Tripathi, C.,K.,M., & Bihari, V., 2008. Production, Optimization and Purification of An Antifungal Compound From *Streptomyces capoaemus* MTCC 8123. *Med.Chem Res*. **17**: 94-102

- [19] Sutrisno, R.B. 1986. TLC (Thin Layer Chromatography). Department of Farmasi, Universitas Pancasila : Jakarta
- [20] Schlatter, D., Fubuh, A., Xiao, K., Hernandez, D., Hobbie, S., & Kinkel, L., 2009. Resource Amendments Influence Density and Competitive Phenotypes of *Streptomyces* in soil. *Microb Ecol.* **57**: 413-420. DOI 10.1007/s00248-008-9433-4
- [21] Ueki, M., Kusumoto, A., Hanafi, M., Shibata, K., Tanaka, T., & Taniguchi, M., 1997. UK-3A, a Novel Antifungal Antibiotic from *Streptomyces* sp. 517-02: Fermentation, Isolation, Structural Elucidation and Biological Properties. *The Journal of Antibiotics* **50** (7): 551-555.
- [22] Worang, R.L. 2003. Endophytic fungi as Antibiotic Producing. *Tesis*. Department of Biology, Universitas Gadjah Mada : Yogyakarta.