

# The Gut-Brain Connection: Exploring the Influence of the Microbiome on Colon Cancer Development and Treatment

Uday Raj Sharma<sup>1\*</sup>, Sushavan Roy<sup>1</sup>, Manjunatha P M<sup>1</sup>, Gurubasavaraja Swamy P M<sup>2</sup>, Suresh Janadri<sup>1</sup>, Surendra Vada<sup>1</sup>, Haribabu. T<sup>1</sup>, Nageena Taj<sup>1</sup>, Gayathri S V<sup>1</sup>

{[udayraj@acharya.ac.in](mailto:udayraj@acharya.ac.in)<sup>1</sup>, [sushavan241@gmail.com](mailto:sushavan241@gmail.com)<sup>1</sup>, [manjunathpm@acharya.ac.in](mailto:manjunathpm@acharya.ac.in)<sup>1</sup>, [gurubasavarajaswamy@gmail.com](mailto:gurubasavarajaswamy@gmail.com)<sup>2</sup>, [sureshjanadri@acharya.ac.in](mailto:sureshjanadri@acharya.ac.in)<sup>1</sup>, [surendrav@acharya.ac.in](mailto:surendrav@acharya.ac.in)<sup>1</sup>, [haribabu@acharya.ac.in](mailto:haribabu@acharya.ac.in)<sup>1</sup>, [nageenataj@acharya.ac.in](mailto:nageenataj@acharya.ac.in)<sup>1</sup>, [gayathri\\_2296@acharya.ac.in](mailto:gayathri_2296@acharya.ac.in)<sup>1</sup>}

<sup>1</sup>Department of Pharmacology, Acharya and BM Reddy College of Pharmacy, Bengaluru, Karnataka, India

<sup>2</sup>Integrated Drug Discovery Centre, Department of Pharmaceutical Chemistry, Acharya & BM Reddy College of Pharmacy, Bengaluru 560107, Karnataka, India

**Abstract.** The gut-brain axis is an intricate bidirectional communication system between the gastrointestinal tract and the central nervous system that the gut microbiota can commonly influence. Recent studies have shown that the gut microbiome is crucial for colorectal cancer development and progression. The focus is on the close interaction between the gut microbiota and the development of colon cancer by explaining the microbial composition, metabolic byproducts and immune modulation. We also investigated how dysbiosis can promote carcinogenesis through inflammation, genotoxicity and immune evasion. In addition, we consider promising microbiota-targeted therapeutic approaches, such as the use of probiotics and faecal microbiota transplantation, together with the development of biotherapeutics and their ability to improve conventional colon cancer treatments. By elucidating the gut-brain-microbiome interplay, this manuscript aims to provide a comprehensive understanding of its impact on colon cancer, opening new avenues for innovative and effective treatment modalities.

**Keywords:** Gut-brain axis, colon cancer, probiotics, prebiotics, faecal microbiota transplantation, microbiome-based biomarkers.

**Abbreviations:** NF- $\kappa$ B, nuclear factor kappa B; iNOS, nitric oxide synthase; JNK, Jun N-terminal kinase; MNNG, N-methyl-N-nitro-N-nitrosoguanidine; MNU, N-methyl-N-nitrosourea; ILCs, Innate lymphoid cells; PTGS2, prostaglandin-endoperoxide synthase; Th 17, T helper 17.

## 1 Introduction

Colon cancer, or colorectal cancer (CRC), is a global health threat influenced by genetic, environmental, and lifestyle factors, ranking third in prevalence and fourth in cancer-related mortality worldwide [1]. Colorectal cancer ranks third in women and fourth in men globally, with regional variations in prevalence; in 2023, the US reported 106,970 new cases, while worldwide, approximately 1.9 million new cases and 935,000 deaths occur annually, according to the WHO [2]. The gut microbiome, a community of bacteria, viruses, and eukaryotes, supports metabolism, immunity, and neurotransmitter production, notably serotonin, highlighting its role in the gut-brain axis [3]. A healthy microbiome balances pro- and anti-inflammatory functions, preventing excessive inflammation while responding quickly to infections. [4] Age, diet, and environment influence the microbiome, but dysbiosis, linked to

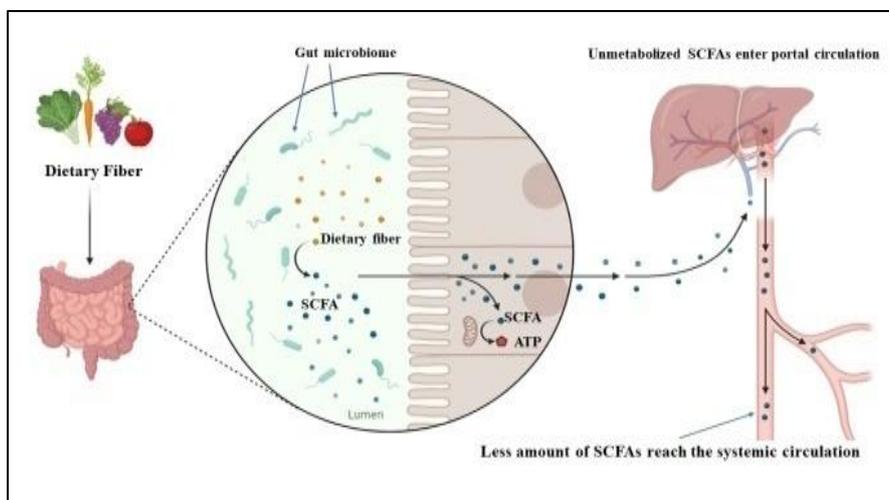
various disorders, causes pronounced changes affecting the gut's transcriptome, proteome, and metabolome[5]. Oxygen may play a role in altering the microbiota composition in individuals with dysbiosis. This concept suggests that increased gut permeability, such as inflammation, leads to increased oxygen availability in the intestinal lumen [6]The gut-brain axis, a bidirectional communication system, influences colon cancer by modulating microbiota composition, inflammation, neurotransmitters, immune responses, and gut barrier integrity, highlighting its role in cancer risk and potential interventions[7].

## **2. The role of microbial metabolites and their effects on colon cancer cells**

Microbial metabolites, small molecules produced in the gut, influence colon health and behaviour, exerting protective or harmful effects based on their nature and context. Some key microbial metabolites and their effects are discussed below.

### **2.1 Short-chain fatty acids (SCFAs)**

Short-chain fatty acids are produced by the gut microbiota from fermentable non-digestible carbohydrates [8]. Two main bacterial groups produce SCFAs: the Bacteroidetes phylum produces acetate and propionate, and *Firmicutes* produces butyrate. These bacteria are also capable of producing additional SCFAs[9]. SCFAs not only affect the colon but can also affect other organs through systemic circulation (Fig.1). Some research studies found that diseases such as diabetes, gastrointestinal distress, atherosclerosis and colorectal cancer can develop due to the influence of SCFAs [10]. Clinical case research has indicated that the concentration of SCFAs is lower than that in the control group, suggesting that there was a reduction in the number of bacterial colonies such as those of *Bifidobacterium* sp., *Roseburia* sp., *Lachnospiraceae*, and other bacteria [11, 12] SCFA receptors are G-Protein coupled receptors (GPCRs) and they are widely distributed throughout the human body. They can affect a variety of cellular pathways. GPR109A, also known as hydroxycarboxylic acid receptor 2 (HCA2, encoded by niacr1), is a receptor found on the apical membranes on the cell surface of macrophages, colonocytes, and adipocytes[13–15]. The role of GPR109A is to release stored fats in situations such as hunger.[16]The only SCFA with anti-tumor action that binds to the receptor GPR109A is butyrate. It can form tight junction proteins and fortify gut integrity by activating the Akt/mTOR signalling pathway in CRC cell line (Caco-2)[17, 18]. Along with downregulating NF-kB activity, it can also enhance the synthesis of cytokines like TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 and decrease the expression of immune regulatory enzymes like COX-2 and iNOS[19]. GPCR43 and GPCR41 regulate appetite, gut function, electrolyte balance, and inflammation, with GPR41 found in enterocytes, renal epithelium, and neurons, while GPR43 is in colonocytes and endocrine cells. Acetate and propionate bind to these receptors, reducing tumor formation and inflammation by phosphorylating JNK and p38, thereby lowering MCP-1 synthesis in response to TNF- $\alpha$ . [20].



**Fig.1.** The microbiota ferments the dietary fiber and produces SCFAs

## 2.2. Conjugated linoleic acids (CLAs)

Conjugated linoleic acids (CLAs) are considered to be beneficial to health and useful food ingredients. A collection of linoleic acid's geometric and positional isomers is known as CLAs. They help in energetic metabolism and function in maintaining body composition [21, 22]. Probiotics such as *Lactobacillus acidophilus*, *Lactobacillus bulgaricus*, *Lactobacillus casei*, and *Lactobacillus plantarum*, etc. use the byproduct of hydrogenation to create CLAs from linoleic acid [23]. It has been demonstrated that CLAs have anti-carcinogenic activity, inhibiting cell proliferation *in-vitro* in human colon cancer cell lines such as HN-29 and Caco-2. It can affect the signalling pathway APC- $\beta$ -catenin-TCF4, which suppresses genes linked to colonocyte multiplication of cells [24]. In addition to this activity, in colon cancer cells (SW 480), CLAs can cause caspase-dependent apoptosis by switching between caspase-3 and caspase-9, raising the level of Annexin V, and reducing the expression of Bcl-2 [25].

## 2.3. Polyamines (PAs)

*Firmicutes* in the gut produce microbial metabolites like cadaverine, putrescine, and spermidine, all of which are polyamines [26, 27]. Host cells can also produce polyamines, which the upper gut can absorb from food [28]. Polyamines, as polycationic compounds, bind to cell's negatively charged components, such as phospholipids in membranes and nucleic acids [29]. The production of siderophores, defence against free radicals and other acids, and stability of the cell wall are all facilitated by polyamines in bacteria [30]. Founded on investigations utilizing cell cultures and animal models. Cancers of the breast, colon, lung, prostate, and skin have also been connected to altered levels of intracellular PAs and changes in their metabolism, in addition to clinical observations [31]. According to a study by Russell and Snyder, cells that are actively dividing, such as rat liver regeneration, chicken embryos, and different tumours, have higher levels of PA synthesis [32]. The enzyme responsible for the rate-limiting process in PA biosynthesis is ornithine decarboxylase (ODC). Elevated ODC activity and, in turn, higher PA concentrations are associated with the development of colorectal cancer (CRC). Probiotic bacteria's ability to change the concentration of PAs in the intestinal lumen is one of their anticancer properties [33]. After feeding the rats a probiotic

cocktail that was enhanced with Bifidobacterium, Lactobacillus, and Streptococcus spp., the rats' PA levels dropped. Additionally, it has been noted that colonocytes exhibit decreased ornithine decarboxylase (ODC) activity[34].

#### **2.4. Polyphenols**

It was found that an increase in the concentration of polyphenols in blood serum decreased the risk of breast cancer and colon cancer [35, 36]. Most of the polyphenols cannot be digested by host cells in the small intestine and enter into the large intestine where bacteria (*Clostridium* sp., *Eubacterium* sp., *Bifidobacterium* sp., *Lactobacillus* sp.) convert those polyphenols into a small number of aromatic compounds, which the body absorbs and circulates throughout the body [37, 38]. Polyphenol metabolites inhibit the cell cycle and induce apoptosis, which has anti-tumor activity [39, 40]. Polyphenols suppress COX-2, modulate cytochrome P450, and induce apoptosis via the mitochondrial pathway. Their phenolic hydroxyl groups enable metal chelation, ROS reduction, and inhibition of NOX and ATPase. By activating the Nrf2/ARE pathway, compounds like resveratrol and curcumin enhance antioxidant defence, while flavonoids and gallic acid mitigate oxidative stress by promoting antioxidant enzymes and preventing lipid peroxidation[41].

#### **2.5. Tryptophan (Trp)**

The necessary amino acid tryptophan (Trp) is a substrate for the bacterial production of vitamin B3, serotonin, and melatonin[42]. Indole lactic acid, indole acetic acid, indole propionic acid, indole acrylic acid, tryptamine, indole acetaldehyde, indole, indolic acid, and indole skatole are the main Trp derivatives [43]. When there is a change in the composition of gut microbiota it can affect the concentration of Trp in plasma [44]. Gut bacteria like *Lactobacillus* sp., *Ruminococcus*navus, and *Clostridium sporogenes* are primarily responsible for the metabolism of tryptophan. Some pathogenic bacteria are also included such as *Escherichia coli*, *Proteus vulgaris*, *Paracolobactrum coliforme*, *Achromobacter liquefaciens*, and *Bacteroides* spp [45, 46]. It has been demonstrated that Trp and the metabolites it produces in bacteria are essential for the development of many types of cancer. Additionally, certain research has indicated that Trp can enhance the malignant characteristics of cancer cells and inhibit the anti-tumor immune response [47]. IDO, a key enzyme in tryptophan metabolism, is overexpressed in cancer and linked to poor prognosis; while Trp-like molecules inhibit IDO as anti-cancer therapy, gut microbiota metabolism may reduce treatment efficacy and impact immune function[48].

#### **2.6. Bile acids**

Cholesterol largely originates in the liver and is the source of primary bile acids, including chenodeoxycholic acid and cholic acid[49]. Bile acids do not directly cause colon cancer in rodents but enhance tumor formation by carcinogens. In humans, high-fat diets increase bile acid release, correlating with CRC risk. Lithocholic and deoxycholic acids induce DNA damage and apoptosis via ROS production. Chronic exposure to deoxycholate may select apoptosis-resistant cells, reducing colon crypt cell apoptosis and promoting cancer development.[50].

#### **2.7. Lipopolysaccharides**

Lipopolysaccharides (LPS) can promote colon carcinogenesis by activating Toll-like receptor 4 (TLR4) signaling pathways, as shown in a TLR4-mediated colon carcinogenesis model [51]. Elevated TLR4 expression in chronic intestinal inflammation induces Cox-2 and PGE2

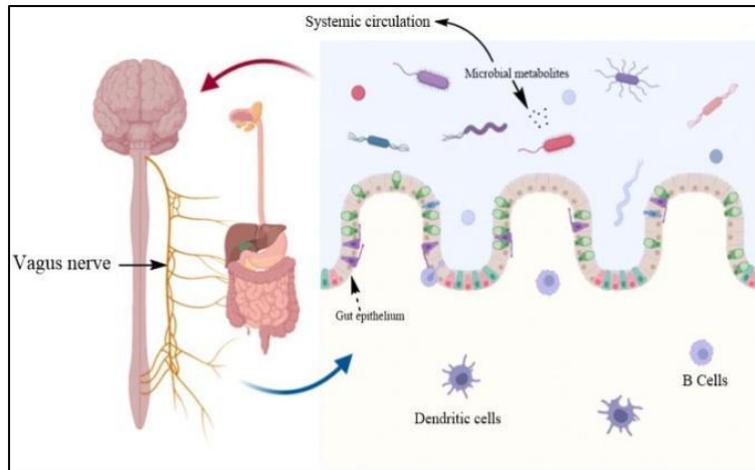
production, promoting colonocyte proliferation, while TLR4 signaling in tumor-associated macrophages and LPS-induced activation contribute to tumor development, immune evasion, DNA damage, and altered apoptosis in colon cancer cells[52].

### **2.8. Aryl hydrocarbon receptor (Ahr) ligands**

The main sources of ligands for the aryl hydrocarbon receptor (AhR), which is present in host immunological and epithelial cells, are microbial metabolites of polyphenols and tryptophan[53–55]. It has been demonstrated that AhR affects gut barrier metabolic balance, mucosal immune response, and cell differentiation. Changes in AhR activity have been shown to induce both intestinal tumors and cell proliferation[56]. When Trp is supplemented during dextran sodium sulphate-induced colitis, AhR activity-induced acute inflammation is reduced, which in turn leads to less cytokines (IL-6, TNF- $\alpha$ , IL-1 $\beta$ ) and chemokines (CCL2, CXCL1, CXCL2) being produced[57]. AhR promotes the growth of innate lymphoid cells (ILCs), which enhances gut integrity and the immune system[58]. ILCs have reduced IL-22 levels and a higher rate of apoptosis when AhR is absent. The onset of CRC corresponds with ILC's synthesis of IL-22. By lowering inflammation and inhibiting the immune system, AhR ligands can support cancer. Through Trp catabolism, cancer cells can create AhR ligands to ward off immune cells[59]. Urolithin A, a microbial metabolite of ellagic acid, exhibits anti-inflammatory effects by reducing PTGS2 and IL-6 expression, though its role in cancer remains under investigation[60].

## **3. The Gut-Brain Axis and Colon Cancer**

There has long been acceptance of the theory that the brain is influenced by the gastrointestinal (GI) system and vice versa. Writers on medicine like Hippocrates, Galen, and Soranus recognized the importance of the stomach and its digestive processes in maintaining morality, physical and mental health, and general well-being[61]. According to Martin et al., and Mayer, the gut-brain axis is a complex communication and control system that connects the gut and brain. It is a bidirectional signalling network made up of neurons, hormones, immune cells, and microbial metabolites[62, 63]. Numerous pathways, including innervated and neuronal ones, as well as small molecule communication systems in the stomach and brain, could be used for communication between the gut and brain [64].The vagus nerve, key to the microbiota-gut-brain axis, links the CNS to gut immunity, where symbiotic bacteria regulate Th17 cell activity and gastrointestinal function[65, 66].Bacteria-derived neurotransmitters and metabolites stimulate the vagus nerve and cross the blood-brain barrier, influencing brain function and systemic responses.Examples of these metabolites are tryptophan derivatives, short-chain fatty acids, and secondary bile acids (Fig. 2). [62, 67–69].Depression and anxiety disrupt the ANS by increasing sympathetic tone, reducing parasympathetic activity, and activating the HPA axis to elevate CRF and cortisol levels[70]. Downstream effects include activation of local inflammatory systems and cytokines, as well as changes in gut flora and secretory activity. These changes may respond to signaling molecules like catecholamines or environmental changes [71].From various research studies, it was found that the progression of colorectal cancer can also occur due to stress, depression and anxiety, these factors change the microenvironment in the gut and due to the change of microenvironment tumor progression takes place [72, 73].



**Fig. 2.** The gut-brain axis communication is bidirectional and mediated by several pathways. This communication is performed through hormonal (hypothalamic-pituitary-adrenal axis), immunological and metabolic pathways (SCFAs and neurotransmitters). Furthermore, the gut microbiota can directly influence the vagus nerve and enteric nervous system, which in turn affects the brain through the gut microbiota, or indirectly through the pathway being affected locally by neuroactive substances (such as noradrenaline, serotonin, dopamine, tryptophan, and short-chain fatty acids).

## 4. Microbiome-Based Interventions for Colon Cancer

Research studies have proven that probiotics have the potential anti-carcinogenic activity, among which colon and gastric cancer cells were commonly studied. A study by Lee et al., showed that the cytoplasmic fractions of *Lactobacillus acidophilus*, *Lactobacillus casei* and *Bifidobacterium longum* have anti-carcinogenic activity in some cancer cell lines [74]. Studies by Russo et al., and Orlando et al., have proved that cytoplasmic extracts of *Lactobacillus rhamnosus* strain GG (LGG) have anti-proliferative action in colon and gastric cancer cells,[75] while *Bifidobacterium adolescentis* SPM0212, a probiotic product, has inhibited the proliferation of three human colon cancer cell lines (HT-29, SW 480 and Caco-2) [76]. In addition to that, Cousin et al., reported that *Propionibacterium freudenreichii* present in fermented milk has enhanced the cytotoxic effect of camptothecin, used as a chemotherapeutic agent for gastric cancer [77].

### 4.1. Mechanistic effect of probiotics for decreasing the chance of colorectal cancer

#### 4.1.1. Modulation of microbiota composition by probiotics

A balanced intestinal microbiota (eubiosis) helps prevent colorectal cancer (CRC), while dysbiosis disrupts this balance, leading to chronic inflammation and carcinogen production, increasing CRC risk[78, 79]. Sobhani *et al.* found significant gut microbiota differences in CRC patients, with higher *Bacteroides*, *Prevotella*, *Clostridium*, and lower *Lactobacillus*, while probiotics can help restore microbial balance by increasing protective lactic acid bacteria[80].*Bacteroides fragilis* is a particular concern because it produces a toxin called fragilysin (BFT). This toxin activates pathways in the body that lead to increased cell growth and inflammation, promoting cancer progression, especially in the advanced stages of CRC

[81]. *Fusobacterium nucleatum*, prevalent in CRC patients, promotes tumor progression and immune evasion, making it a potential CRC biomarker[82]. *Escherichia coli*, especially strains from groups B2 and D, produce harmful toxins that can interfere with cell processes and contribute to chronic inflammatory bowel diseases, increasing the risk of CRC [83]. Other bacteria, such as *Streptococcus gallolyticus* and *Enterococcus faecalis*, are also associated with CRC [84]. Probiotics aid cancer prevention by producing antimicrobial substances like lactic acid, hydrogen peroxide, and bacteriocins, inhibiting harmful bacteria and competing for space and nutrients in the gut[85].

#### **4.1.2 Beneficiation of Probiotics for the Intestinal Epithelial Barrier**

Intestinal barrier dysfunction allows allergen penetration and pathogenic invasion, triggering localized inflammation and impairing epithelial defense, increasing susceptibility to colorectal cancer (CRC)[86]. According to Hsieh et al., certain strains of Bifidobacterium enhance the integrity of the epithelium and shield it from TNF- $\alpha$ -induced disruption[87]. Lactobacillus probiotics strengthen tight junctions (occludin, claudin-3), reduce oxidative stress and inflammation, and help restore intestinal barrier function after colitis[88]. Beneficial bacteria preserve the integrity of tight junction proteins and increase mucus defensins from the goblet cells, which aid in the reconstruction of the epithelial barrier. This process prevents harmful substances and antigens from leaking and enhances this barrier's integrity [86, 89].

#### **4.1.3. Probiotics metabolites against colorectal cancer and as an anti-oxidant**

The probiotic bacteria produce short-chain fatty acids, conjugated linoleic acids, and phenols when they ferment dietary fibre. These compounds have anti-cancer properties that can fight colorectal cancer [78]. Butyric acid, produced by bacteria like *Clostridium* and *Roseburia*, supports gut health by promoting cancer cell apoptosis, regulating colon cell balance, strengthening the intestinal barrier, reducing inflammation, and modulating key proteins involved in apoptosis and antioxidant activity[78, 90]. Probiotic strains of Lactobacillus and Propionibacterium produce conjugated linoleic acid (CLA), which exhibits anti-inflammatory and anti-cancer effects by suppressing eicosanoids and modulating the PPAR- $\gamma$  receptor[91]. Some probiotics like *Lactobacillus* and *Pedicoccus* can even produce antioxidants that may inhibit tumours from forming [92].

#### **4.1.4. Probiotics as an anti-inflammatory agent**

Probiotics support immune function by promoting immune cell growth, regulating responses, inducing antioxidants and anti-inflammatory compounds, and modulating cytokine expression to prevent immune-related diseases[93]. Probiotics interact with TLRs, modulate IL-8 and TNF production, inhibit NF- $\kappa$ B activation, and influence PPAR- $\gamma$  and MAPK pathways, supporting immune regulation and anti-cancer responses. Lactobacillus strains modulate immune responses by influencing cytokine secretion, enhancing cellular immunity, producing SCFAs like butyrate and propionate, and stimulating T and B lymphocytes to support anti-cancer activity and intestinal protection[94]. Probiotics and their metabolic byproducts, including fatty acids, CLA, and phenols, support normal cell development, reduce DNA damage, inhibit cancer cell growth, and induce apoptosis, aiding in colorectal cancer prevention and treatment.

## 5. Fecal microbiota transplantation as a Therapeutic approach in Colon cancer treatment

Fecal microbiota transplantation (FMT) restores gut microbiota, treating *Clostridium difficile* infections, IBD, hematological malignancies, and antibiotic-resistant bacterial colonization, including resistant strains like *Klebsiella pneumoniae*, ESBL+ *Escherichia coli*, and *Pseudomonas aeruginosa*, with 75% achieving complete bacterial decolonization and 80% showing partial reduction, even in neutropenic patients[95,96]. FMT is gaining interest as a potential colorectal cancer treatment by reducing inflammation, slowing cell proliferation, and counteracting cancer-related pathways, but further research is needed to confirm its efficacy. [97,98]. Figure 3 represents the process of fecal microbiota transplantation from a healthy donor to a patient through endoscopy or oral capsules.

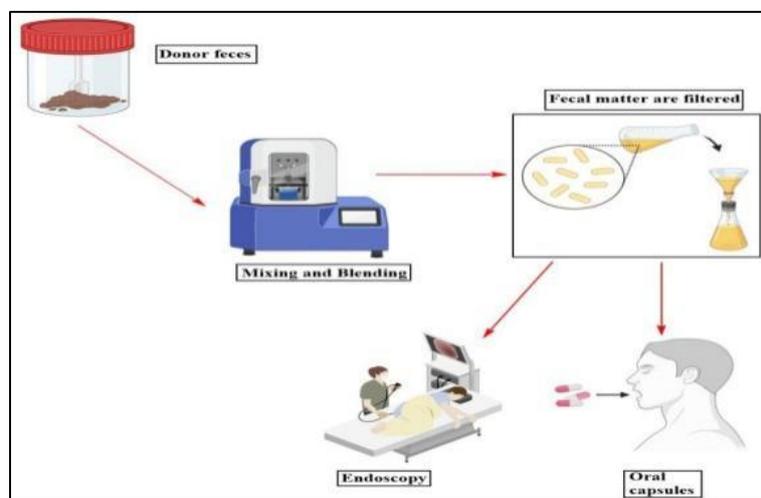


Fig. 3. Schematic diagram of fecal microbiota transplantation

## 6. Conclusions and Future Perspectives

Colon cancer risk is closely tied to the gut microbiome, with dysbiosis potentially promoting tumor growth. Factors like diet, lifestyle, and stress impact this microbiome, influencing cancer progression through microbial metabolites and the gut-brain axis. Innovations such as probiotics, prebiotics, and fecal microbiota transplantation (FMT) show promise for prevention and treatment, while microbial biomarkers may enable early detection. However, challenges like individual microbiome variability, ethical concerns, and limited understanding of mechanisms remain. Future research into microbial pathways, engineered probiotics, and integrated therapies could lead to personalized strategies, improving outcomes and addressing the multifaceted nature of colon cancer.

## References

- [1] Gupta D, Tomar S. Introduction to the Comprehensive Overview of Colon Cancer: Unveiling the Epidemiological Trends, Risk Factors, Molecular Mechanisms, and Therapeutic Interventions. *PEXACY International Journal of Pharmaceutical Science*. 2023 Sep 3;2(8):169-90.
- [2] Ide Souza JS, Reitz LK, Copetti CL, Moreno YM, Vieira FG, Di Pietro PF. Lower Adherence to Lifestyle Recommendations of the World Cancer Research Fund/American Institute for Cancer Research (2018) Is Associated with Decreased Overall 10-Year Survival in Women with Breast Cancer. *Nutrients*. 2025 Mar 12;17(6):1001.
- [3] O'Mahony SM, Clarke G, Borre YE, Dinan TG, Cryan JF (2015) Serotonin, tryptophan metabolism and the brain-gut-microbiome axis. *Behavioural Brain Research* 277:32–48
- [4] Tizard IR, Jones SW (2018) The Microbiota Regulates Immunity and Immunologic Diseases in Dogs and Cats. *Veterinary Clinics of North America: Small Animal Practice* 48:307–322
- [5] Zeng MY, Inohara N, Nuñez G (2017) Mechanisms of inflammation-driven bacterial dysbiosis in the gut. *Mucosal Immunol* 10:18–26
- [6] Rigottier-Gois L (2013) Dysbiosis in inflammatory bowel diseases: the oxygen hypothesis. *ISME J* 7:1256–1261
- [7] Schledwitz A, Xie G, Raufman J-P (2021) Exploiting unique features of the gut-brain interface to combat gastrointestinal cancer. *Journal of Clinical Investigation*. <https://doi.org/10.1172/JCI143776>
- [8] Miller TL, Wolin MJ (1996) Pathways of acetate, propionate, and butyrate formation by the human fecal microbial flora. *Appl Environ Microbiol* 62:1589–1592
- [9] Levy M, Thaiss CA, Elinav E (2016) Metabolites: messengers between the microbiota and the immune system. *Genes Dev* 30:1589–1597
- [10] Parada Venegas D, De la Fuente MK, Landskron G, González MJ, Quera R, Dijkstra G, Harmsen HJM, Faber KN, Hermoso MA (2019) Short Chain Fatty Acids (SCFAs)-Mediated Gut Epithelial and Immune Regulation and Its Relevance for Inflammatory Bowel Diseases. *Front Immunol*. <https://doi.org/10.3389/fimmu.2019.00277>
- [11] Wang T, Cai G, Qiu Y, Fei N, Zhang M, Pang X, Jia W, Cai S, Zhao L (2012) Structural segregation of gut microbiota between colorectal cancer patients and healthy volunteers. *ISME J* 6:320–329
- [12] Yusuf F, Adewiah S, Syam AF, Fatchiyah F (2019) Altered profile of gut microbiota and the level short chain fatty acids in colorectal cancer patients. *J Phys Conf Ser* 1146:012037
- [13] J, Faller D, Spanjaard R (2003) Short-Chain Fatty Acid Inhibitors of Histone Deacetylases: Promising Anticancer Therapeutics? *Curr Cancer Drug Targets* 3:219–236
- [14] Kasubuchi M, Hasegawa S, Hiramatsu T, Ichimura A, Kimura I (2015) Dietary Gut Microbial Metabolites, Short-chain Fatty Acids, and Host Metabolic Regulation. *Nutrients* 7:2839–2849

- [15] Thangaraju M, Cresci GA, Liu K, et al (2009) GPR109A Is a G-protein–Coupled Receptor for the Bacterial Fermentation Product Butyrate and Functions as a Tumor Suppressor in Colon. *Cancer Res* 69:2826–2832
- [16] Blad CC, Ahmed K, IJzermanAdP, Offermanns S (2011) Biological and Pharmacological Roles of HCA Receptors. pp 219–250
- [17] Singh N, Gurav A, Sivaprakasam S, et al (2014) Activation of Gpr109a, Receptor for Niacin and the Commensal Metabolite Butyrate, Suppresses Colonic Inflammation and Carcinogenesis. *Immunity* 40:128–139
- [18] Feng W, Wu Y, Chen G, Fu S, Li B, Huang B, Wang D, Wang W, Liu J (2018) Sodium Butyrate Attenuates Diarrhea in Weaned Piglets and Promotes Tight Junction Protein Expression in Colon in a GPR109A-Dependent Manner. *Cellular Physiology and Biochemistry* 47:1617–1629
- [19] Chen G, Ran X, Li B, Li Y, He D, Huang B, Fu S, Liu J, Wang W (2018) Sodium Butyrate Inhibits Inflammation and Maintains Epithelium Barrier Integrity in a TNBS-induced Inflammatory Bowel Disease Mice Model. *EBioMedicine* 30:317–325
- [20] Kuwahara A, Kuwahara Y, Inui T, Marunaka Y (2018) Regulation of Ion Transport in the Intestine by Free Fatty Acid Receptor 2 and 3: Possible Involvement of the Diffuse Chemosensory System. *Int J Mol Sci* 19:735
- [21] Kobayashi M, Mikami D, Kimura H, Kamiyama K, Morikawa Y, Yokoi S, Kasuno K, Takahashi N, Taniguchi T, Iwano M (2017) Short-chain fatty acids, GPR41 and GPR43 ligands, inhibit TNF- $\alpha$ -induced MCP-1 expression by modulating p38 and JNK signaling pathways in human renal cortical epithelial cells. *BiochemBiophys Res Commun* 486:499–505
- [22] Benjamin S, Spener F. Conjugated linoleic acids as functional food: an insight into their health benefits. *Nutrition & Metabolism*. 2009 Dec;6:1-3.
- [23] Lehn TE, da Silva MR, Camacho A, Marcadenti A, Lehn AM (2015) A review on effects of conjugated linoleic fatty acid (CLA) upon body composition and energetic metabolism. *J Int Soc Sports Nutr*. <https://doi.org/10.1186/s12970-015-0097-4>
- [24] Ewaschuk JB, Walker JW, Diaz H, Madsen KL (2006) Bioproduction of Conjugated Linoleic Acid by Probiotic Bacteria Occurs In Vitro and In Vivo in Mice. *J Nutr* 136:1483–1487
- [25] Lampen A, Leifheit M, Voss J, Nau H (2005) Molecular and cellular effects of cis-9, trans-11-conjugated linoleic acid in enterocytes: Effects on proliferation, differentiation, and gene expression. *Biochimica et Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids* 1735:30–40
- [26] Miller A, Stanton C, Devery R (2002) Cis 9, trans 11- and trans 10, cis 12-conjugated linoleic acid isomers induce apoptosis in cultured SW480 cells. *Anticancer Res* 22:3879–87
- [27] Hanfrey CC, Pearson BM, Hazeldine S, Lee J, Gaskin DJ, Woster PM, Phillips MA, Michael AJ (2011) Alternative Spermidine Biosynthetic Route Is Critical for Growth of *Campylobacter jejuni* and Is

the Dominant Polyamine Pathway in Human Gut Microbiota. *Journal of Biological Chemistry* 286:43301–43312

[28] Matsumoto M, Benno Y (2007) The Relationship between Microbiota and Polyamine Concentration in the Human Intestine: A Pilot Study. *Microbiol Immunol* 51:25–35

[29] Minois N, Carmona-Gutierrez D, Madeo F (2011) Polyamines in aging and disease. *Aging* 3:716–732

[30] Schuber F (1989) Influence of polyamines on membrane functions. *Biochemical Journal* 260:1–10

[31] Field AM, Rowatt E, Williams RJP (1989) The interaction of cations with lipopolysaccharide from *Escherichia coli* C as shown by measurement of binding constants and aggregation reactions. *Biochemical Journal* 263:695–702

[32] Nowotarski SL, Woster PM, Casero RA (2013) Polyamines and cancer: implications for chemotherapy and chemoprevention. *Expert Rev Mol Med* 15:e3

[33] Russell D, Snyder SH (1968) Amine synthesis in rapidly growing tissues: ornithine decarboxylase activity in regenerating rat liver, chick embryo, and various tumors. *Proceedings of the National Academy of Sciences* 60:1420–1427

[34] Giardiello FM, Hamilton SR, Hylind LM, Yang VW, Tamez P, Casero RA (1997) Ornithine decarboxylase and polyamines in familial adenomatous polyposis. *Cancer Res* 57:199–201

[35] Linsalata M, Russo F, Berloco P, Valentini AM, Caruso ML, De Simone C, Barone M, Polimeno L, Di Leo A (2005) Effects of probiotic bacteria (VSL#3) on the polyamine biosynthesis and cell proliferation of normal colonic mucosa of rats. *In Vivo* 19:989–95

[36] Pietinen P, Stumpf K, Männistö S, Kataja V, Uusitupa M, Adlercreutz H (2001) Serum enterolactone and risk of breast cancer: a case-control study in eastern Finland. *Cancer Epidemiol Biomarkers Prev* 10:339–44

[37] Kuijsten A, Arts ICW, Hollman PCH, van't Veer P, Kampman E (2006) Plasma Enterolignans Are Associated with Lower Colorectal Adenoma Risk. *Cancer Epidemiology, Biomarkers & Prevention* 15:1132–1136

[38] Espín JC, González-Sarriás A, Tomás-Barberán FA (2017) The gut microbiota: A key factor in the therapeutic effects of (poly)phenols. *Biochem Pharmacol* 139:82–93

[39] Selma M V., Espín JC, Tomás-Barberán FA (2009) Interaction between Phenolics and Gut Microbiota: Role in Human Health. *J Agric Food Chem* 57:6485–6501

[40] Aires V, Limagne E, Cotte AK, Latruffe N, Ghiringhelli F, Delmas D (2013) Resveratrol metabolites inhibit human metastatic colon cancer cells progression and synergize with chemotherapeutic drugs to induce cell death. *Mol Nutr Food Res* 57:1170–1181

[41] Stanisławska I, Granica S, Piwowarski J, Szawkało J, Wiązecki K, Czarnocki Z, Kiss A (2019) The Activity of Urolithin A and M4 Valerolactone, Colonic Microbiota Metabolites of Polyphenols, in a Prostate Cancer In Vitro Model. *Planta Med* 85:118–125

- [42] Karlsson PC, Huss U, Jenner A, Halliwell B, Bohlin L, Rafter JJ (2005) Human Fecal Water Inhibits COX-2 in Colonic HT-29 Cells: Role of Phenolic Compounds. *J Nutr* 135:2343–2349
- [43] Long J, Guan P, Hu X, et al (2021) Natural Polyphenols as Targeted Modulators in Colon Cancer: Molecular Mechanisms and Applications. *Front Immunol*. <https://doi.org/10.3389/fimmu.2021.635484>
- [44] Slominski A, Semak I, Pisarchik A, Sweatman T, Szczesniewski A, Wortsman J (2002) Conversion of L-tryptophan to serotonin and melatonin in human melanoma cells. *FEBS Lett* 511:102–106
- [45] Smith EA, Macfarlane GT (1997) Formation of Phenolic and Indolic Compounds by Anaerobic Bacteria in the Human Large Intestine. *MicrobEcol* 33:180–188
- [46] Clarke G, Stilling RM, Kennedy PJ, Stanton C, Cryan JF, Dinan TG (2014) Minireview: Gut Microbiota: The Neglected Endocrine Organ. *Molecular Endocrinology* 28:1221–1238
- [47] Gao J, Xu K, Liu H, Liu G, Bai M, Peng C, Li T, Yin Y (2018) Impact of the Gut Microbiota on Intestinal Immunity Mediated by Tryptophan Metabolism. *Front Cell Infect Microbiol*. <https://doi.org/10.3389/fcimb.2018.00013>
- [48] Keszthelyi D, Troost FJ, Masclee AAM (2009) Understanding the role of tryptophan and serotonin metabolism in gastrointestinal function. *Neurogastroenterology & Motility* 21:1239–1249
- [49] Pilotte L, Larrieu P, Stroobant V, et al (2012) Reversal of tumoral immune resistance by inhibition of tryptophan 2,3-dioxygenase. *Proceedings of the National Academy of Sciences* 109:2497–2502
- [50] Günther J, Däbritz J, Wirthgen E (2019) Limitations and Off-Target Effects of Tryptophan-Related IDO Inhibitors in Cancer Treatment. *Front Immunol*. <https://doi.org/10.3389/fimmu.2019.01801>
- [51] Björkhem I, Eggertsen G (2001) Genes involved in initial steps of bile acid synthesis. *Curr Opin Lipidol* 12:97–103
- [52] Bernstein H, Bernstein C, Payne CM, Dvorakova K, Garewal H (2005) Bile acids as carcinogens in human gastrointestinal cancers. *Mutation Research/Reviews in Mutation Research* 589:47–65
- [53] Fukata M, Chen A, Klepper A, et al (2006) Cox-2 Is Regulated by Toll-Like Receptor-4 (TLR4) Signaling: Role in Proliferation and Apoptosis in the Intestine. *Gastroenterology* 131:862–877
- [54] Fukata M, Abreu MT (2008) Role of Toll-like receptors in gastrointestinal malignancies. *Oncogene* 27:234–243
- [55] Hubbard TD, Murray IA, Bisson WH, Lahoti TS, Gowda K, Amin SG, Patterson AD, Perdev GH (2015) Adaptation of the human aryl hydrocarbon receptor to sense microbiota-derived indoles. *Sci Rep* 5:12689
- [56] Koper JEB, Loonen LMP, Wells JM, Troise AD, Capuano E, Fogliano V (2019) Polyphenols and Tryptophan Metabolites Activate the Aryl Hydrocarbon Receptor in an in vitro Model of Colonic Fermentation. *Mol Nutr Food Res*. <https://doi.org/10.1002/mnfr.201800722>
- [57] Rooks MG, Garrett WS (2016) Gut microbiota, metabolites and host immunity. *Nat Rev Immunol* 16:341–352

- [58] Metidji A, Omenetti S, Crotta S, Li Y, Nye E, Ross E, Li V, Maradana MR, Schiering C, Stockinger B (2018) The Environmental Sensor AHR Protects from Inflammatory Damage by Maintaining Intestinal Stem Cell Homeostasis and Barrier Integrity. *Immunity* 49:353-362.e5
- [59] Islam J, Sato S, Watanabe K, et al (2017) Dietary tryptophan alleviates dextran sodium sulfate-induced colitis through aryl hydrocarbon receptor in mice. *J NutrBiochem* 42:43–50
- [60] Kiss EA, Vonarbourg C, Kopfmann S, Hobeika E, Finke D, Esser C, Diefenbach A (2011) Natural Aryl Hydrocarbon Receptor Ligands Control Organogenesis of Intestinal Lymphoid Follicles. *Science* (1979) 334:1561–1565
- [61] Murray IA, Patterson AD, Perdew GH (2014) Aryl hydrocarbon receptor ligands in cancer: friend and foe. *Nat Rev Cancer* 14:801–814
- [62] Muku G, Murray I, Espin J, Perdew G (2018) Urolithin A Is a Dietary Microbiota-Derived Human Aryl Hydrocarbon Receptor Antagonist. *Metabolites* 8:86
- [63] LeBlanc RL. Tolstoy's Body: Diet, Desire, and Denial. In *Cultures of the Abdomen: Diet, Digestion, and Fat in the Modern World 2005* (pp. 147-166). New York: Palgrave Macmillan US.
- [64] Martin CR, Osadchiy V, Kalani A, Mayer EA (2018) The Brain-Gut-Microbiome Axis. *Cell Mol Gastroenterol Hepatol* 6:133–148
- [65] Mayer EA (2011) Gut feelings: the emerging biology of gut–brain communication. *Nat Rev Neurosci* 12:453–466
- [66] Keightley PC, Koloski NA, Talley NJ (2015) Pathways in gut-brain communication: Evidence for distinct gut-to-brain and brain-to-gut syndromes. *Australian & New Zealand Journal of Psychiatry* 49:207–214
- [67] Bonaz B, Bazin T, Pellissier S (2018) The Vagus Nerve at the Interface of the Microbiota-Gut-Brain Axis. *Front Neurosci*. <https://doi.org/10.3389/fnins.2018.00049>
- [68] Chow J, Mazmanian SK (2009) Getting the Bugs out of the Immune System: Do Bacterial Microbiota “Fix” Intestinal T Cell Responses? *Cell Host Microbe* 5:8–12
- [69] Cryan JF, Dinan TG (2012) Mind-altering microorganisms: the impact of the gut microbiota on brain and behaviour. *Nat Rev Neurosci* 13:701–712
- [70] Cussotto S, Sandhu K V., Dinan TG, Cryan JF (2018) The Neuroendocrinology of the Microbiota-Gut-Brain Axis: A Behavioural Perspective. *Front Neuroendocrinol* 51:80–101
- [71] Forsythe P, Kunze WA (2013) Voices from within: gut microbes and the CNS. *Cellular and Molecular Life Sciences* 70:55–69
- [72] Coss-Adame E, Rao SSC (2014) Brain and Gut Interactions in Irritable Bowel Syndrome: New Paradigms and New Understandings. *Curr Gastroenterol Rep* 16:379
- [73] Bailey MT, Dowd SE, Galley JD, Hufnagle AR, Allen RG, Lyte M (2011) Exposure to a social stressor alters the structure of the intestinal microbiota: Implications for stressor-induced immunomodulation. *Brain Behav Immun* 25:397–407

- [74] Carson TL, Byrd DA, Smith KS, Carter D, Gomez M, Abaskaron M, Little RB, Holmes SN, van Der Pol WJ, Lefkowitz EJ, Morrow CD. A case-control study of the association between the gut microbiota and colorectal cancer: exploring the roles of diet, stress, and race. *Gut Pathogens*. 2024 Mar 11;16(1):13.
- [75] Di Y-Z, Han B-S, Di J-M, Liu W-Y, Tang Q (2019) Role of the brain-gut axis in gastrointestinal cancer. *World J Clin Cases* 7:1554–1570
- [76] Lee J-W, Shin J-G, Kim EH, Kang HE, Yim IB, Kim JY, Joo H-G, Woo HJ (2004) Immunomodulatory and antitumor effects in vivo by the cytoplasmic fraction of *Lactobacillus casei* and *Bifidobacterium longum*. *vetsci.org* 41–48
- [77] Orlando A, Refolo MG, Messa C, Amati L, Lavermicocca P, Guerra V, Russo F (2012) Antiproliferative and proapoptotic effects of viable or heat-killed *Lactobacillus paracasei* IMPC2.1 and *Lactobacillus rhamnosus* GG in HGC-27 gastric and DLD-1 colon cell lines. *Nutr Cancer* 64:1103–1111
- [78] Kim KJ, Kim Y, Lee D, Kim D, Cho J, Yang J, Chung M, Kim K, Ha N (2008) Inhibition of proliferation in colon cancer cell lines and harmful enzyme activity of colon bacteria by *Bifidobacterium adolescentis* SPM0212. *SpringerY Kim, D Lee, D Kim, J Cho, J Yang, M Chung, K Kim, N HaArchives of pharmacal research, 2008•Springer* 31:468–473
- [79] Cousin FJ, Jouan-Lanhouet S, Dimanche-Boitrel MT, Corcos L, Jan G (2012) Milk fermented by *propionibacteriumfreudenreichii* induces apoptosis of HGT-1 human gastric cancer cells. *PLoS One*. <https://doi.org/10.1371/JOURNAL.PONE.0031892>
- [80] dos Reis SA, da Conceição LL, Siqueira NP, Rosa DD, da Silva LL, Peluzio M do CG (2017) Review of the mechanisms of probiotic actions in the prevention of colorectal cancer. *Nutrition Research* 37:1–19
- [81] Akin H, Tözün N (2014) Diet, Microbiota, and Colorectal Cancer. *J Clin Gastroenterol* 48:S67–S69
- [82] Sobhani I, Tap J, Roudot-Thoraval F, Roperch JP, Letulle S, Langella P, Corthier G, Van Nhieu JT, Furet JP (2011) Microbial Dysbiosis in Colorectal Cancer (CRC) Patients. *PLoS One* 6:e16393
- [83] Boleij A, Hechenbleikner EM, Goodwin AC, et al (2015) The *Bacteroides fragilis* Toxin Gene Is Prevalent in the Colon Mucosa of Colorectal Cancer Patients. *Clinical Infectious Diseases* 60:208–215
- [84] Kostic AD, Gevers D, Pedamallu CS, et al (2012) Genomic analysis identifies association of *Fusobacterium* with colorectal carcinoma. *Genome Res* 22:292–298
- [85] Buc E, Dubois D, Sauvanet P, Raisch J, Delmas J, Darfeuille-Michaud A, Pezet D, Bonnet R (2013) High Prevalence of Mucosa-Associated *E. coli* Producing Cyclomodulin and Genotoxin in Colon Cancer. *PLoS One* 8:e56964
- [86] Ambalam P, Raman M, Purama RK, Doble M (2016) Probiotics, prebiotics and colorectal cancer prevention. *Best Pract Res Clin Gastroenterol* 30:119–131

- [87] Gosai V, ambalam P, Raman M, Kothari CR, Kothari RK, Vyas BRM, Sheth NR (2011) Protective effect of *Lactobacillus rhamnosus* 231 against N-Methyl-N'-nitro-N-nitrosoguanidine in animal model. *Gut Microbes* 2:319–325
- [88] Kahouli I, Tomaro-Duchesneau C, Prakash S (2013) Probiotics in colorectal cancer (CRC) with emphasis on mechanisms of action and current perspectives. *J Med Microbiol* 62:1107–1123
- [89] Hsieh C-Y, Osaka T, Moriyama E, Date Y, Kikuchi J, Tsuneda S (2015) Strengthening of the intestinal epithelial tight junction by *Bifidobacterium bifidum*. *Wiley Online Library* CY Hsieh, T Osaka, E Moriyama, Y Date, J Kikuchi, S Tsuneda *Physiological reports*, 2015•Wiley Online Library. <https://doi.org/10.14814/phy2.12327>
- [90] Cui Y, Liu L, Dou X, Wang C, Zhang W, Gao K, Oncotarget JL-, 2017 undefined *Lactobacillus reuteri* ZJ617 maintains intestinal integrity via regulating tight junction, autophagy and apoptosis in mice challenged with lipopolysaccharide. *ncbi.nlm.nih.gov* Y Cui, L Liu, X Dou, C Wang, W Zhang, K Gao, J Liu, H Wang *Oncotarget*, 2017•ncbi.nlm.nih.gov
- [91] Konieczna C, ... AO-S-AS, 2018 undefined *Lactobacillus* spp. belonging to the Casei group display a variety of adhesins. *food.actapol.net* CKonieczna, A Olejnik-Schmidt, MT Schmidt *Acta Scientiarum Polonorum Technologia Alimentaria*, 2018•food.actapol.net. <https://doi.org/10.17306/J.AFS.2018.0538>
- [92] Czajkowska A, Experimental BS-A in H and, 2018 undefined Short chain fatty acids (SCFA), the products of gut bacteria metabolism and their role in the host. *phmd.pl* Czajkowska, B Szponar *Advances in Hygiene and Experimental Medicine*, 2018•phmd.pl. <https://doi.org/10.5604/01.3001.0011.6468>
- [93] Bassaganya-Riera J, Viladomiu M, Pedragosa M, et al (2012) Probiotic Bacteria Produce Conjugated Linoleic Acid Locally in the Gut That Targets Macrophage PPAR  $\gamma$  to Suppress Colitis. *PLoS One* 7:e31238
- [94] Ewaschuk J, Walker J, Diaz H, nutrition KM-TJ of, 2006 undefined Bioproduction of conjugated linoleic acid by probiotic bacteria occurs in vitro and in vivo in mice. Elsevier
- [95] Cristofori F, Dargenio VN, Dargenio C, Miniello VL, Barone M, Francavilla R (2021) Anti-Inflammatory and Immunomodulatory Effects of Probiotics in Gut Inflammation: A Door to the Body. *Front Immunol*. <https://doi.org/10.3389/FIMMU.2021.578386/FULL>
- [96] Molska M, Reguła J (2019) Potential Mechanisms of Probiotics Action in the Prevention and Treatment of Colorectal Cancer. *Nutrients* 11:2453
- [97] Chen D, Wu J, Jin D, Wang B, Cao H (2019) Fecal microbiota transplantation in cancer management: Current status and perspectives. *Int J Cancer* 145:2021–2031
- [98] Bilinski J, Grzesiowski P, Sorensen N, et al (2017) Fecal Microbiota Transplantation in Patients With Blood Disorders Inhibits Gut Colonization With Antibiotic-Resistant Bacteria: Results of a Prospective, Single-Center Study. *Clinical Infectious Diseases* 65:364–370